Synthesis, Chemical Reactivity, and Cytotoxicity of 2-Bis(alkoxycarbonyl)methyliden-1-azabicyclo[3.1.0]hexane Systems Related to Antitumor Antibiotic Carzinophilin A<sup>†</sup>

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Enantiomeric pairs of the title compounds were synthesized starting from (S)-and (R)-pyroglutamic acid. They were found to be susceptible to nucleophilic ring opening of aziridine moieties and to exhibit weak *in vitro* cytotoxicity.

Carzinophilin A (Cz-A) (1) is an antitumor antibiotic isolated from *Streptomyces sahachiroi* by Hata *et al.* in 1954.<sup>1)</sup> While the structure of 1 has been revised several times by plural groups over 30 years,<sup>2)</sup> Armstrong *et al.* reported in 1991 that <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of 1 were superimposable on those of azinomycin B.<sup>3)</sup> The latter compound has been isolated as an antitumor antibiotic from other *Streptomyces* species and carries a characteristic 2-methyliden-1-azabicyclo[3.1.0]hexane ring system.<sup>4)</sup> Since 1 is known as one of the strand cross-rinking compounds for DNA,<sup>5)</sup> it is anticipated that, as shown in 2, the antitumor activity of 1 is probably due to cleavage of the strained aziridine ring by attack of a nucleotide which might effect denaturation of DNA.<sup>6)</sup>

Since Armstrong *et al.* recently reported the synthesis of the model compound of Cz-A [(Z)-isomer] (3),<sup>7</sup>) we wish to report here our own results in this area, culminating in successful preparation of the enantiomeric pair

<sup>†</sup>Dedicated to Professor Emeritus Osamu Simamura of The University of Tokyo on the occasion of his 80th birthday.

of novel 2-bis(alkoxycarbonyl)methyliden-1-azabicyclo[3.1.0]hexane system (4 and ent-4) related to 1. These bicyclic compounds (4 and ent-4) were originally designed by expecting that their aziridine moieties could be cleaved in a similar manner to that proposed for 2. As expected, 4 and ent-4 were found to undergo facile nucleophilic ring opening of their aziridine moieties and, moreover, to exhibit weak in vitro cytotoxicity.

The synthesis of 4 and ent-4 commences from optically pure (S)- and (R)-5-hydroxymethyl-2-pyrrolidone (5 and ent-5) obtainable from (S)- and (R)-pyroglutamic acid according to the reported procedure. For convenience, the synthetic route employing 5 is shown in Scheme 1. Thus, the hydroxyl group of 5 was first protected in a form of TBDPS ether. The 2-pyrrolidone moiety of the silyl ether was converted into a thiolactam by using Lawesson's reagent, yielding the pyrrolidin-2-thione (6). Treatment of 6 with diethyl or dibenzyl bromomalonate followed by the addition of aqueous KHCO3 solution resulted in the formation of the 2-bis(alkoxycarbonyl)methylidenpyrrolidines  $(7a,b)^{10}$  in good yields. After the TBDPS ethers of 7a,b were cleaved by the usual method, the generated alcohols were converted into the tosylates (8a,b), which were further transformed into the bromides (9a,b) and the iodides (10a,b), respectively.

With completion of the synthesis of three types of the reaction substrates (8-10), the intramolecular aziridine formation producing a 2-bis(alkoxycarbonyl)methyliden-1-azabicyclo[3.1.0]hexane system was attempted. It was found that treatment of 8a with potassium hydride in THF at room temperature for 15 min smoothly effected the construction of aziridine ring, giving rise to 4a in 80% yield based on the <sup>1</sup>H-NMR spectrum of the crude reaction product (crude yield). Since 4a was quite unstable under acidic conditions (vide infra), its isolation

$$\begin{array}{c} a, b \\ 97\% \\ (2 \text{ steps}) \end{array}$$

$$\begin{array}{c} a, b \\ 97\% \\ (2 \text{ steps}) \end{array}$$

$$\begin{array}{c} a : 97\% \\ b : 88\% \end{array}$$

$$\begin{array}{c} TBDPSO \end{array}$$

$$\begin{array}{c} A : 90\% \\ b : 92\% \\ TBDPSO \end{array}$$

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$$\begin{array}{c} CO_2R \\ D : 2 \text{ steps} \end{array}$$

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a) TBDPSCl, ImH., DMF, rt, 12 h b) Lawesson's reagent, toluene, reflux, 20 min c) diethyl bromomalonate or dibenzyl bromomalonate,  $CH_2Cl_2$ , rt, 12 h, then aq. KHCO<sub>3</sub>, 3-4 h d) TBAF, THF, rt, 1-2 h e) TsCl, Py.,  $CH_2Cl_2$ , rt, 1-2 d f) Bu<sub>4</sub>NBr,  $CH_3CN$ , reflux, 3-4 h g) NaI, acetone, reflux, 15-18 h h) KH, THF, rt, 15 min i) KH, THF, rt, 2 h

was able to be achieved only by quick Florisil column chromatography, affording a pure sample of 4a as an oil,  $[\alpha]_D^{20}$  +104° (c 0.750, CHCl<sub>3</sub>), <sup>11)</sup> in 34% yield (isolated yield). This rather low yield might reflect that 4a decomposes during purification process. While 9a also cyclized to 4a in 70% crude yield, its isolation in a pure state turned out to be fruitless due to low separation efficiency of 4a from 9a by Florisil column chromatography. The cyclization of 8b to 4b took place in a very low yield under the same conditions as employed for 8a. However, desired  $4b^{12}$ ) was obtained in 17% isolated yield by using 9b as a reaction substrate. Long range couplings (1.0 and 0.9 Hz, respectively) between signals of  $C_3$ -H $\alpha$  and  $C_6$ -H $\alpha$  observed in the  $^1$ H-NMR spectra of 4a,b as well as NOE experiments carried out on 4a obviously established the structures of 4a,b as depicted. It is notable that treatment of 10a,b under the same conditions as employed for 8a,b and 9a,b gave the dimeric products (11a,b) in 57% and 70% yields, respectively, after 2 hrs' reactions, although production of a small amounts of 4a,b was always detected by TLC analysis of the reaction mixture. Accordingly, the formation of 11a,b might be explained by coupling of produced 4a,b with the metaloamides of unreacted 10a,b followed by intramolecular cyclization. Following the same procedures as described above, ent-4a,  $[\alpha]_D^{20}$ -93.0° (c 0.800, CHCl<sub>3</sub>), and ent- $4b^{12}$ ) were prepared from ent-5.

4a 
$$\frac{a, b, \text{ or c}}{a: 80\%}$$

$$\frac{\text{CO}_2\text{Et}}{\text{b: } 74\%}$$

$$\frac{\text{c: } 62\%}{\text{c: } 62\%}$$

$$\frac{a, b, \text{ or c}}{\text{a: } 80\%}$$

$$\frac{\text{CO}_2\text{Et}}{\text{b: } X = \text{OMe}}$$

$$\frac{\text{b: } X = \text{OMe}}{\text{b: } X = \text{OAc}}$$

$$\frac{\text{c: } X = \text{SPh}}{\text{c: } 62\%}$$

a) MeOH, rt, 7 d b) AcOH, THF, rt, 3 h c) PhSH, Et<sub>3</sub>N, THF, rt, 3 h

## Scheme 2.

With 4a,b and ent-4a,b in hand, the chemical reactivity of 2-bis(alkoxycarbonyl)methyliden-1-azabicyclo-[3.1.0]hexane system was next studied by employing 4a as a representative substrate. Thus, a pure sample of 4a was found to be stable in CDCl<sub>3</sub> at 0 °C for a week. However, its slow decomposition was observed in MeOH at room temperature, producing the 5-methoxymethylpyrrolidine (12a). The T<sub>1/2</sub> value for the decomposition of 4a was estimated as ca. 2.5 days. Acetic acid reacted with 4a in THF to afford the 5-acetoxymethylpyrrolidine (12b) smoothly. Even under weakly basic conditions, for example, when 4a was treated with thiophenol in the presence of triethylamine in THF at room temperature, cleavage of the aziridine ring occurred to give the 5-phenylthiomethylpyrrolidine (12c). In these experiments, no formation of the 5-substituted piperidine derivatives (13a-c) was observed. The structures of 12a-c were definitely verified by their <sup>1</sup>H-NMR and mass spectra. Highly regioselective formation of 12a-c obviously suggests that nucleophilic ring opening of the 2-bis(alkoxycarbonyl)methyliden-1-azabicyclo[3.1.0]hexane systems takes place solely at their C<sub>6</sub>-positions as initially expected.

Finally, *in vitro* cytotoxicity of 4a,b and *ent*-4a,b was examined. Thus, when 4a,b and *ent*-4a,b were subjected to *in vitro* cytotoxicity assay against P388 murine leukemia, the following very weak cytotoxicity was observed for these compounds [IC<sub>50</sub> (µg/mL): 0.74 (4a), 11.6 (4b), 3.1 (*ent*-4a), and 7.6 (*ent*-4b)].

As described above, we have succeeded in preparing 4a,b bearing a novel 2-bis(alkoxycarbonyl)-methyliden-1-azabicyclo[3.1.0]hexane system related to 1 and in exploring their chemical reactivity and *in vitro* cytotoxicity. Based on these results, synthetic studies aiming at the second generation of the model compound of 1 are in progress in these laboratories.

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- 6) In their studies on the isolation of azinomycins, Yokoi *et al.* reported that the naphthalene moiety of 1, which lacks the peptide part corresponding to 2, exibited no cytotoxicity.<sup>4)</sup>
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- 11) <sup>1</sup>H-NMR spectral data of **4a,b** (CDCl<sub>3</sub>) are as follows; **4a** δ 1.20, 1.26 (each 3H, t, J= 7.1 Hz, CH<sub>3</sub>×2), 1.61 (1H, d, J= 5.1 Hz, C<sub>6</sub>H<sub>α</sub>), 2.14 (2H, m, C<sub>4</sub>H<sub>2</sub>), 2.40 (1H, dd, J= 1.0 and 4.3 Hz, C<sub>6</sub>H<sub>β</sub>), 2.61 (1H, ddt, J= 1.0, 17.9, and 4.3 Hz, C<sub>3</sub>H<sub>α</sub>), 2.88 (1H, m, C<sub>5</sub>H), 3.25 (1H, ddd, J= 6.4, 8.7, and 17.9 Hz, C<sub>3</sub>H<sub>β</sub>), 4.11 (1H, q, J= 7.1 Hz, CH<sub>2</sub>CH<sub>3</sub>), 4.22 and 4.31 (each 1H, dq, J= 10.8 and 7.1 Hz, CH<sub>2</sub>CH<sub>3</sub>); **4b** δ 1.67 (1H, d, J= 4.3 Hz, C<sub>6</sub>H<sub>α</sub>), 2.19 (2H, m, C<sub>4</sub>H<sub>2</sub>), 2.42 (1H, dd, J= 0.9 and 5.0 Hz, C<sub>6</sub>H<sub>β</sub>), 2.70 (1H, ddt, J= 0.9, 20.1, and 8.0 Hz, C<sub>3</sub>H<sub>α</sub>), 2.98 (1H, m, C<sub>5</sub>H), 3.33 (1H, ddd, J= 6.9, 7.2, and 20.1 Hz, C<sub>3</sub>H<sub>β</sub>), 5.15 (2H, s, CH<sub>2</sub>Ph), 5.21, 5.35 (each 1H, d, J= 12.3 Hz, CH<sub>2</sub>Ph), and 7.20-7.40 (10H, m, aromatic protons). In the spectrum of **4a**, NOEs were observed between the signals of C<sub>3</sub>-H<sub>α</sub> and C<sub>6</sub>-H<sub>β</sub> (2.6%), C<sub>5</sub>-H and C<sub>6</sub>-H<sub>β</sub> (4.9%), and C<sub>6</sub>-H<sub>α</sub> and C<sub>6</sub>-H<sub>β</sub> (23%).
- 12) Optical rotation values of 4b and ent-4b were not measured because of their low yields.
- 13) Representative <sup>1</sup>H-NMR (CDCl<sub>3</sub>) and mass (EI) spectral data are as follows; **12a** δ 1.67 (1H, m, C<sub>4</sub>H), 2.10 (1H, m, C<sub>4</sub>H), 3.05-3.50 (7H, m, C<sub>3</sub>H<sub>2</sub>, C<sub>5</sub>CH<sub>2</sub>, CH<sub>3</sub>O), and 4.03 (1H, m, C<sub>5</sub>H), m/z 271 (rel. int., 15, M+) and 180 (100, M-EtO-MeOCH<sub>2</sub>-H+); **12b** δ 1.67 (1H, dddd, *J*= 6.2, 6.8, 9.3, and 13.1 Hz, C<sub>3</sub>H), 2.03 (3H, s, CH<sub>3</sub>CO), 2.10 (1H, dddd, *J*= 5.8, 8.0, 9.4, and 13.1 Hz, C<sub>3</sub>H), 3.02 (1H, ddd, *J*= 6.9, 9.4, and 18.5 Hz, C<sub>4</sub>H), 3.13 (1H, ddd, *J*= 5.8, 9.5, and 18.5 Hz, C<sub>4</sub>H), 3.93 (1H, dd *J*= 7.6 and 11.2 Hz, C<sub>2</sub>CHH), 4.04 (1H, m, C<sub>2</sub>H), and 4.16 (1H, dd, *J*= 3.9 and 11.2 Hz, C<sub>2</sub>CHH), m/z 299 (rel. int., 2.7, M+), 226 (4.1, M-AcOCH<sub>2</sub>+), and 180 (100, M-EtO-AcOCH<sub>2</sub>-H+); **12c** δ 1.78 (1H, dddd, *J*= 6.0, 6.8, 9.4, and 12.9 Hz, C<sub>4</sub>H), 2.21 (1H, dddd, *J*= 5.7, 7.7, 9.1, and 12.9 Hz, C<sub>4</sub>H), 2.96 (1H, dd *J*= 7.3 and 13.4 Hz, C<sub>5</sub>CHH), 3.05 (1H, ddd, *J*= 6.8, 9.5, and 18.5 Hz, C<sub>3</sub>H), 3.07 (1H, dd *J*= 5.8 and 13.4 Hz, C<sub>5</sub>CHH), 3.23 (1H, ddd, *J*= 5.7, 9.4, and 18.5 Hz, C<sub>3</sub>H), and 3.98 (1H, br quint, *J*= 6.0 Hz, C<sub>5</sub>H), m/z 349 (rel. int., 2.1, M+), 226 (11, M-PhSCH<sub>2</sub>+), 180 (100, M-EtO-PhSCH<sub>2</sub>-H+).